

Fungal contamination of Azadi Teaching Hospital and Hevi Paediatric Hospital Environments, Duhok, Iraq

Shilan S. Saleem¹, Wasan M. Alnakshabandie¹, Asia A. M. Saadullah²

¹ College of Health Sciences, University of Duhok, Duhok, Iraq

² Biology Department, College of Science, University of Duhok, Duhok, Iraq
asiasaadullah73@yahoo.com

Abstract

This survey study was undertaken to find out the prevalence and distribution of fungi occupying the residential surfaces in Azadi Teaching Hospital and Hevi Paediatric Hospital in Duhok province, Iraq. Three hundred samples were collected from randomly selected areas in laboratories, wards, consultation rooms, operating theatres and hospital kitchen by swabs. These samples were cultured on two selective fungal media (PDA and SDA) and also samples from air were cultured on PDA in order to determine the genus or species of the agents in the samples. The results found microorganisms that were identified included forty eight isolates in addition to non identified yeasts and sterile mycelium also detected. From 300 samples collected from period January to May 2016, molds comprised 60% and yeasts comprised 40%. All fungal isolates were identified depending on the morphological and microscopic examinations as well as biochemical tests. The environmental contaminations in Azadi Teaching and Hevi Paediatric Hospitals with fungi (*Aspergillus*) were the highest fungal isolates whereas *Candida* sp. was the highest yeast isolate.

Key words: Fungi, *Aspergillus*, *Candida*, Environmental swabs.

Introduction

Fungi are ubiquitous in the environment but they rarely cause symptoms in human beings due to the effective defence mechanisms, such as the cell-mediated response. Different fungi causing infections, some are commonly occurring infections while the others are rare¹. Fungal contamination in healthcare facilities has been the subject of numerous studies. These have shown that high percentages of hospital infections are caused by fungi, such as *Candida* sp. and various species of *Aspergillus*, *Penicillium*, and *Cladosporium*². Hospitals are considered the favourite places for the emergence of a wide variety of opportunistic and pathogenic fungi; these fungi are ubiquitous and can be acquired from host surroundings or are components of normal endogenous flora. However, any decreasing in the host organism's defensive ability due to cancer, HIV infection³⁻⁴, medical therapy and organ transplantation may lead to uncontrolled multiplication of fungi and consequent onset of infection. Among kidney transplant recipients, 11% of infections were of fungal origin. In 47% of such cases, infection occurred during the first three months after transplantation⁵. After solid organ transplantation, 80% of fungal infections involved *Candida* and *Aspergillus* species. Among immunocompromised patients in general, the incidence of invasive Aspergillosis and other fungal diseases have increased during the last few decades which remains a serious complication and could be fatal⁶.

Airborne fungal conidia are inhaled by everyone, because their concentration is high in the air⁷, this may be the reason why nosocomial acquired infections and community acquired infections quite often develop in immunocompromised and in immunocompetent people⁸.

The aim of this study was to determine the level of fungal contamination in Azadi Teaching Hospital and Hevi Paediatric Hospital in Duhok province.

Material and methods

Samples collection

From first January to end of May 2016, 300 samples were taken using sterile transport media swabs from various environments including operating theatres, intensive care, laboratories, outpatient departments, kitchens, and other working environments like nurses' offices, and refectories in both hospitals.

Culturing laboratory:

Swabs cultivated directly on the media culture appropriate (Sabouraud dextrose agar, Malt extract agar and Potato dextrose agar) and exposure of PDA plates to air. The dishes were incubated at 25 and 37 °C for 7-35 days. By the end of this period, the cultures examined for any growth and then sub-culture of the dishes that have shown positive results⁹.

Identification of Fungi :

During incubation period, different fungal colonies were subjected to macroscopic and microscopic to observe their growth, mycelium nature and structure of hyphae. Filamentous fungal growth - as mold and yeast- that grow on SDA, MEA and PDA were sub-cultured on separate SDA culture plates. One plate was incubated at 37° C, and the remaining was incubated at 25° C¹⁰. Pure culture growth of each mold and yeast colony was examined under magnification for their microscopic structures and cross identified by using mycological keys manuals¹¹⁻¹⁷.

Biochemical tests used for identification of *candida* species³⁸.

Frequency Percentage:

The percentage frequency of species isolated was calculated by applying the following formula.

$$\text{Percentage frequency} = \frac{\text{Number of isolates of the same species}}{\text{Total number of isolates of all species}} \times 100$$

Occurrence Percentage: The percentage for the Occurrence of each species isolated was calculated according to the following equation.

$$\text{Percentage of Occurrence} = \frac{\text{Number of samples that appeared to show one type}}{\text{Total number of samples}} \times 100$$

Results and Discussion

Table-1 shows number of swabs, percentages of contamination in both Azadi Teaching and Hevi Paediatric Hospitals. As shown in the table the percentage of positive contamination in Azadi Hospital is more than the percentage of contamination in Hevi Paediatric Hospital, this could be due to the high number of patients occupied in this hospital.

Table 1 - The number of swabs and percentages of contamination.

Hospitals	Azadi	Hevi	Total
Swabs	190	110	300
Positive	121	63	184
%	63.6	57.2	61.33
Negative	69	47	116
%	36.3	42.7	38.66

Table-2 shows the genera of fungi isolated from both hospitals using two media PDA and SDA by swab spreading method and exposure of plates of PDA to air for 15 minutes.

The results shown thirty two species of fungi in addition to sterile mycelium and non identified yeast have been demonstrated. Thirty species of fungi were isolated on PDA using swab method; the highest

number obtained was *Aspergillus* sp. (264), *Penicillium* sp. (125), *Ulocladium* sp. (86), *Cladosporium* sp. (75), followed by *Candida* sp. (69) and *Rhizopus* (35).

Twenty seven species of fungi were isolated on PDA plates exposure to air, the highest number obtained was *Aspergillus* sp. (71), *Ulocladium* sp. (39), *Penicillium* sp. (30), followed by *Cladosporium* sp. (25) and *Candida* sp. (17).

Twenty two species of fungi were isolated on SDA using swab method; the highest number obtained was *Aspergillus* sp. (55), *Penicillium* sp. (39), *Candida* sp. (19), Dermatophytes (16) followed by *Ulocladium* sp. (13), and *Rhizopus* sp. (11).

The difference between the numbers of fungi isolated on both media was expected because PDA is a selective medium for luxuriant growth of yeasts and molds. Potato infusion encourages mold sporulation and pigment production even in some dermatophytes¹⁸.

PDA enhances the production of reproductive structures (conidia) and colony colour when other media fail to do so; it is often recommended for slide culture or for inducing an isolate to exhibit a characteristic pigment¹⁹. Because of low pH SDA is useful for the cultivation of fungi particularly those associated with skin infections²⁰.

Table (2): Isolated Fungi by different culture media.

No.	Fungal isolates	Potato Dextrose Agar swab	Potato Dextrose Agar air	Saboroud Dextrose Agar	Total
1	<i>Absidia</i> sp.	7	2	5	14
2	<i>Acremonium</i> sp.	12	0	0	12
3	<i>Alternaria</i> sp.	4	9	0	13
4	<i>Aspergillus</i> sp.	138	71	55	264
5	<i>Aurobasidium</i> sp.	0	1	0	1
6	<i>Candida</i> sp.	33	17	19	69
7	<i>Chaetomium</i> sp.	1	0	1	2
8	<i>Chrysosporium</i> sp.	1	0	0	1
9	<i>Cladosporium</i> sp.	41	25	9	75
10	<i>Curvularia</i> sp.	1	0	3	4
11	<i>Dermatophytes fungi</i>	10	7	16	33
12	<i>Epicoccum</i> sp.	1	1	0	2
13	<i>Emericella</i> sp.	2	1	0	3
14	<i>Exophiala</i> sp.	12	6	2	20
15	<i>Fusarium</i> sp.	10	6	3	19
16	<i>Gliaocladium</i> sp.	3	0	3	6
17	<i>Geotrichum</i> sp.	8	8	5	21
18	<i>Gymnoascus</i> sp.	0	1	1	2
19	<i>Monilia</i> sp.	3	1	6	10
20	<i>Microascus</i> sp.	2	3	0	5
21	<i>Mucor</i> sp.	8	8	6	22
22	<i>Neoscytalidium</i> sp.	15	3	9	27
23	<i>Pacilomyces</i> sp.	1	3	0	4
24	<i>Penicillium</i> sp.	56	30	39	125
25	<i>Phialophora</i> sp.	4	2	0	6
26	<i>Phoma</i> sp.	2	0	0	2
27	<i>Rhizopus</i> sp.	17	7	11	35
28	<i>Stachybotrys</i> sp.	12	4	7	23
29	<i>Sterile mycelium</i> (pink)	22	19	16	57
30	<i>Talaromyces</i> sp.	0	1	0	1
31	<i>Trichoderma</i> sp.	4	1	1	6
32	<i>Ulocladium</i> sp.	34	39	13	86
33	<i>Verticillium</i> sp.	1	0	0	1
34	<i>Yeasts black</i> (non identified)	0	2	27	29
Total		255	275	257	1000

Table-3 shows fungal species, frequency and occurrence percentages of the genera and species of fungi isolated during the study. From 300 swabs and air samples (28) genera and 31 species in addition to non identified yeasts and sterile mycelium has been identified.

This table shows the high frequency percentage of fungi this indicates widespread of fungi in both hospitals and may be due to appropriate environmental conditions specially temperature and humidity.

The most common fungi collected were, *Aspergillus*, *Penicillium*, *Candida*, *Cladosporium*, *Alternaria*, *Rhizopus*, *Mucor*, yeast and *Fusarium* species

Aspergillus was represented by 12 species and showed the widest diversity among all recovered genera. *A. niger*, *A. flavus* and *A. fumigatus* showed the high frequency and occurrence percentages because these species are ubiquitous and the major airborne contaminant fungi.

Some species can cause infection in humans and animals. The most common pathogenic species are *A. fumigatus* and *A. flavus*. *A. fumigatus* infections are primary pulmonary infections includes Aspergillosis, Pneumonia, Asthma and Allergy²¹. *A. flavus* cause infection of the external ear and Aspergillosis²².

Overberger *et al.* (1995) found that 70 - 80% of the fungi in hospitals air were *Aspergillus* sp. Our study showed similar distribution of the mold and yeast species²³.

Penicillium was second in the number of species isolated and was represented by ten species. These species are usually found in indoor environment and cause infections to humans and animals.

This study has shown *Candida* fungi were isolated from indoor environment in all selected units. As a rule, candidiasis is an endogenous infection; however, exogenous infections are also possible. Numerous

studies have shown evidence that *Candida* fungi are detected in the indoor air at e.g. surgical, haematological and obstetric wards; they are potential and major source of infections, especially in risk group patients²⁴⁻²⁶.

Another yeast-like organism like *Geotrichum candidum* and *Aurobasidium* are opportunistic organism that causes pulmonary and systemic infections in human and animals.

Alternaria species are emerging as opportunistic pathogens. A case of Phaeohyphomycosis caused by *A. infectoria* in a renal transplant recipient has been reported²⁷.

Chrysosporium species have been isolated in this study and some were reported to cause superficial skin infection.

Microascus species are considered as potentially pathogenic to human beings and are frequently isolated from floor dusts²⁸⁻³⁰.

Fusarium species may cause a range of opportunistic infections in humans. For example nails (onychomycosis) and in the cornea (keratomycosis or mycotic keratitis)

Zygomycetes fungi isolated in this study include *Absidia*, *Mucor* and *Rhizopus* that cause brain infections via nose³¹.

Many fungi isolated from both hospitals like *A. flavus*, *A. ochraceus*, *A. parasiticus*, *A. fumigatus*, *Emericella nidulans*, *Fusarium* and *Stachybotrys* secrete secondary metabolites which harm human and animals during inhalation and are known as mycotoxins³²⁻³³. Dermatophytes also were isolated in this study including *Microsporum*, *Epidermphyto* and *Trichophyton*; however, some people are at greater risk than others. The fungus takes advantage of skin of those patients with reduced immune capacity.

Our results in this study are in line with many studies³⁴⁻³⁷.

Table (3) The species and genus fungal isolates and the frequency and occurrence percentages

No.	Fungi	Frequency percentage	Occurrence percentage
1	<i>Absida sp.</i>	2.0	1.0
2	<i>Alternaria alternate</i> (Fr.) Keissl		2.0
3	<i>Aureobasidium sp.</i>	3.3	1.2
4	<i>A.carbonarius</i> (Bainier) Thom	1.3	0.8
5	<i>A.flavus</i> Link	21.0	13.0
6	<i>A.foetidus</i>	1.5	
7	<i>A.fumigatus</i> Fresen	5.34	4.0
8	<i>A.italicum</i>	0.0	0.24
9	<i>A.japonicus</i> Saito	1.50	0.22
10	<i>A.niger</i> Tiegh.nom.cons.	63.4	50
11	<i>A.niveus</i> Blochwitz		1.0
12	<i>A.ochraceus</i> K.Wilh	4.0	1.0
13	<i>A.parasiticus</i> Speare	3.5	2.5
14	<i>A.vadensis</i> Samson ,devries,Frisvad&visser		0.8
15	<i>A.terreus</i>	0.33	0.2
16	<i>Penicillium aurantiogresum</i> Dierckx	0.4	0.2
17	<i>P.chrysogenum</i> Thom		2.0
18	<i>Penicillium citrinum</i>	7.0	5.0
19	<i>P.expansum</i>		1.0
20	<i>P.glabrum</i> (Wehmer) Westling	1.33	0.7
21	<i>P.greaseofulvum</i>	1.0	
22	<i>P.italicum</i>	1.0	1.0
23	<i>P.naglivoience</i> Laxa	2.0	1.0
24	<i>P.oxalicum</i> Currie & Thom	2.66	2.0
25	<i>P.verrocsum</i> Dierckx		2.33
26	<i>Candida sp.</i>	24.0	18.0
27	<i>Cladosporium cladosporoides</i> (Fresen.) G.A. de Vries	6.33	4.0
28	<i>Cladosporium herborum</i> (Pers.) Link	3.0	2.0
29	<i>Curvularia sp.</i>	3.0	1.0
30	<i>Emericella nidulans</i> (Eidam) Vuill	0.33	0.1
31	<i>Epidermphyton sp.</i>	1.24	0.18
32	<i>Exophiala sp.</i>	0.04	0.02
33	<i>Phoma glomerata</i> (Corda) Wollenw.&Hochapfel	0.2	0.19
34	<i>Fusarium oxysoprum</i> Schlecht	0.17	
35	<i>Geotrichum candidum</i> Link ex Fr.	2.0	1.0
36	<i>Gliocladium sp.</i>	2.0	
37	<i>Gymnoascus sp.</i>		0.46
38	<i>Microsporium sp.</i>	0.16	0.02
39	<i>Monilia sp.</i>	2.0	0.46
40	<i>Microascus sp.</i>	1.0	0.33
41	<i>Mucor circinelloides</i> Tiegh	4.0	
42	<i>Paecilomyces varioti</i> Bainier	1.6	0.33
43	<i>Rhizopus stolonifer</i> (Ehrenb.) Vuill.	1.0	
44	<i>Rhodotorulla sp.</i>	1.0	0.33
45	<i>Stachybotrys sp.</i>	3.0	
46	<i>Sterile mycelium</i>	11.0	9.0
47	<i>Trichoderma sp.</i>	1.33	0,44
48	<i>Trichophyton sp.</i>	1.33	0.48
49	<i>Ulocladium atrum</i>	0.46	0.33
50	<i>Yeast non identified</i>	1.0	0.64

Conclusion

In conclusion, the present study revealed the presence of potentially pathogenic and toxigenic fungi in both hospitals in Duhok province. Strict hygienic measures

should therefore be undertaken to control the spread of these fungi in the environment and to avoid mycotic infection among the patients and staff alike. Check air conditioning systems for standing moisture and clean or replace as necessary.

References

- 1- Ryan, K.J. and Ray, C.G. (2010). *Sherris Medical Microbiology*. 5th ed. The McGraw-Hill com. USA.
- 2- Perfect, J.R. and Casadevall, A. (2006). Fungal molecular pathogenesis: what can it do and why do we need it?. p. 3–11. In: *Molecular principles of fungal pathogenesis*. Heitman, J., (ed.). ASM Press. Washington DC.
- 3- Khan, M.S.A., Ahmad, I., Aqil, F., Owais, M., Shahid, M. and Musarrat, J. (2010). Virulence and pathogenicity of fungal pathogens with special reference to *Candida albicans*. p: 21-45. In: Ahmad, E.A. (ed.). *Combating fungal infections*. Springer-Verlag Berlin Heidelberg.
- 4- Klein, B.S. and Tebbets, B. (2007). Dimorphism and virulence in fungi. *Curr. Opin. Microbiol.* 10:314–319.
- 5- Richardson, M.D. (2005). Changing patterns and trends in systemic fungal infections. *J. Antimicrob. Chemo. Ther.*, 56, pp: 5-11.
- 6- Kontoyiannis DP, Bodey GP. Invasive aspergillosis in 2002: an update. *Eur J Clin Microbiol Infect Dis* 2002; 21:161-172.
- 7- Buczynska, A., Cyprowski, M., Piotrowska, M. and Szadkowska- Stanczyk, J. (2007). Indoor moulds: results of the environmental study in office rooms. *Med. Pr.* 58: 521–525.
- 8- Warris, A., Klaassen, C.H., Meis, J.F., De Ruiter, M.T., De Valk, H.A., Abrahamsen, T.G., Gaustad, P. and Verweij, P.E. (2003). Molecular epidemiology of *Aspergillus fumigatus* isolates recovered from water, air, and patients shows two clusters of genetically distinct strains. *J. Clin. Microbiol.*, 41, pp: 4101–4106.
- 9- Grossi P, Farina C, Fiocchi R, Dalla Gasperina D. Prevalence and outcome of invasive fungal infections in (1963) thoracic organ transplant recipients: a multicenter retrospective study. Italian Study Group of Fungal Infections in Thoracic Organ Transplant Recipients. *Transplantation*(2000); 70:112-116.
- 10- Koneman, E.W; Allin, S.D; Janad, W.M; Schreckenberger, P.C and Winn, W.C (1997). *Color Atlas and Textbook of Diagnostic Microbiology*, 5 th ed. USA: J. B. Lippincott Company..
- 11- Matsushima , T. (1975). *Icones Micofungorum A Matsushima leetorum*. Koba, Japan. 413 p.
- 12- Domsch, K.H.; Gams, W.; Anderson, T.H. (1980). "Compendium of soil fungi". Academic Press. London.
- 13- Horre, Y" (1980) Ascospore Ornamentation and its application to the taxonomic re-evaluation in *Emericella* . *Trans . Mycol. Soc. Japan* " 21:4g3_493 .
- 14- Kreger–Van, R. (1984). *The yeasts: a taxonomie study*. 3rd ed. Elsevier Science Publishers B. V. Amsterdam, Netherlands.
- 15- Ellis, M. B. and Ellis, J. P. (1985). *Microfungi on Land Plants: An Identification Handbook* (1st ed.). Macmillan Pub Co.
- 16- Sivanesan , A. (1987) Graminicolous species of *Bipolaris Curvulaira*, *Drechslera*, *Exerohilum* and their teleomorphs. CAB. International. Kew, Surrey, U.K.
- 17- Hoog , G.S", de and Guarro , J. (1995) " Atlas of clinical fungi Centraalbureureuschele culture Barn and Universitat Rovir Ivirgili Spain720pp.
- 18- Japanese Pharmacopoeia, (2007). the MHLW Ministerial Notification Sixteenth Edition.
- 19- Larone, D. (2011). *Medically Important Fungi: A Guide to Identification*. Ed. (5). Washington, DC: ASM Press.
- 20- Murray, P.R., Baron, E.J., Jorgensen, J.H., Tenover, F.C., and Tenover, R.H. (2003). *Manual of Clinical Microbiology* , 8th ed., American Society for Microbiology, Washington, DC.
- 21- Kim, O.G., Horg, S.C., Kim, H. J. Chi, J. G. Han, M.h., Chosi, K.S. and Han, D.H. (1993) Cerebral aspergillosis in immunologically competent patients. *Surg. Neurol* . 4: 326-331 .
- 22- Drakos, P.E., Nagler, A., Orr, R., Naparstek, E., Kapelushink, J., Engelhard, D., Rahay, G. and Slavin, S. (1993). Invasive fungal sinusitis in patients undergoing bone marrow transplantation *Transpl*" 12:208.
- 23- Overberger, P A; Wadowsky, R M and Schaper, M M. (1995). Evaluation of airborne particulates and fungi during hospital renovation. *American Indoor Hygiene Assessment Journal* 56: 706-712.
- 24- Kao, AS; Brandt, ME; Pruitt, WR; Conn, LA; Perkins, BA. Stephens, DS; Baughman, WS; Reingold AL; Rothrock, GA; Pfaller, MA; Pinner, RW and Hajjeh, RA. (1999).The epidemiology of candidemia in two United States cities: results of apopulation-based active surveillance. *Cli Infect Dis*; 29: 1164.
- 25- Vanden Bergh, MF.; Verwij, PE and Voss, A. (1999). Epidemiology of nosocomial fungal infections: invasive aspergillosis and the environment. *Diag Microbiol Infect Dis*, 34: 221-7.FA.
- 26- Pegues, D. ; Lasker, B; McNeil, M.; Hamm, P.; Lundal, J. and Kubak, B. (2002).Cluster of cases on invasive aspergillosis in a transplant intensive care unit: evidence of person – to – person airborne transmission. *Clin Infect Dis*, 34(3): 412-6.
- 27- Halaby, I.; Boots, H.; Vermulen, A.; Van derv en, A.; Beguin, H.; Van Hooft, H., Jacobs, J. (2001). phaeohyphomycoses caused by *Alternaria infectoria* in a renal transplant recipient. *J. Clin. Microbiol.*, 39,1952-1955.
- 28- Abdel-Hafez, A.M. (1991). The incidence of dermatophytes, keratinophilic and saprophytic fungi in a playground of schools and house dusts in upper Egypt. *Cryptogamie Mycol.*, 12,305-314.
- 29- Gughani, H.C. (2000). Non-dermatophytic filamentous keratinophilic fungi and their role in human infection . In "Biology of dermatophytes and

other keratinophilic fungi" (eds.) Kushwaha, R.K.S and Guarro, J. Revista Iberoamericana Micologia, Spain. Pp 109-114.

30- Abdullah, S.K.; Al-Musa, A.A. (2000). The incidence of keratinophilic and actidione resistant fungi in the floor dust of residential houses in Basrah. *Basrah J.Sci.*, 18,45-54.

31- Ferry, A.P. and Abedi, S" (1993) Diagnosis and management of Rhion_ orbitocerbral mucomycesis *Ophthalmology* 90: 1096_1104 "

32- Croft, W" A. Jarvis, R. B. and Yatawara, C.S. (1986) " Airborne outbreak of trichothecene toxieosis. *Atmos Environ*.2A: 549-552".

33- Miller, J.D. (1992) Fungi as a contaminants in indoor air . *Atoms. Environ* " 26:2163-2172 .

34- Abdullah, S.K. and Al-Ani, S.A.(2003). Air borne fungal flora in indoor environment of two hospitals in Basrah city .*Iraqi J.Biology*,3: 60-67.

35- Perdelli, F ; M. L. Cristina.; M. Sartini.; Spagnolo A. M., (2006). Fungal Contamination in Hospital Environments. *infection control and hospital epidemiology* january, vol. 27, no. 1.

36- Mohammed , Sadiq R. AL-Jibouri and Mona H. (2015). Isolation and identification of fungi from two hospitals in Baghdad city and effect of disinfectants on some fungi. *Iraqi Journal of Science*, 2015, Vol 56, No.1C, pp: 673-682.

37- Abbas Soleymani MD, Seyed Ali Asghar Sefidgar C.P, Hamze Sharifi MS (2015). Species Diversity of Keratinophilic Fungi in Various Soil Type of Babol Medical University's Hospitals' Yard. *International Journal of Applied Science and Technology*. Vol. 5, No. 3; June 2015.

38-Aslanzadeh, J. and G.D. Roberts, 1991. Direct microscopic examination of clinical specimens for the laboratory diagnosis of fungal infections. *Clin. Microbiol. Newsl.*, 13: 185-192.

التلوث الفطري في بيئة مستشفى آزادي التعليمي ومستشفى هيفي للأطفال

محافظة دهوك، العراق

شيلان صدقي سليم¹ ، وسن مدحت يوسف¹ ، اسيا عبد الحميد محمد²

¹كلية علوم الصحية ، جامعة دهوك ، دهوك ، العراق

²قسم البايولوجي ، كلية العلوم ، جامعة دهوك ، دهوك ، العراق

asiasaadullah73@yahoo.com

الملخص

أجريت هذه الدراسة لغرض معرفة مدى أنتشار الفطريات المستوطنة والمنتشرة في بيئة كل من مستشفى آزادي التعليمي ومستشفى هيفي للأطفال في محافظة دهوك، حيث تم جمع 300 عينة بطريقة القطن المعقم (السواب) من مناطق مختلفة وعشوائية شملت: المختبرات، الاقسام، غرف الاستشارية، غرف العمليات والمطابخ وزرعت هذه العينات على نوعين من الاوساط الفطرية الاختيارية (وسط بطاطا دكستروز اكار، وسط سابرويد دكستروز اكار) كما زرعت العينات المأخوذة من الهواء على وسط بطاطا دكستروز اكار لتحديد نوع وجنس الفطريات الموجودة في هذه العينات. وخلال هذه الدراسة تم تحديد 48 نوع فطري بالإضافة الى الخمائر والماسيليوم. وجمعت هذه العينات (300 عينة) في فترة كانون الثاني الى ايار سنة 2016 حيث بلغت الاعفان نسبة 60% وبلغت الخمائر 40%. وتم تشخيص جميع الفطريات بالأعتماد على الفحوصات المظهرية والمجهريية بالإضافة الى اجراء الفحوصات البايوكيميائية. وأظهرت الرشاشيات (*Aspergillus*) من بين الاعفان كأعلى نسبة من الملوثات الفطرية في كل من بيئة مستشفى آزادي التعليمي ومستشفى هيفي للأطفال كمان اظهرت المبيضات (*Candida sp*) وانواعها أعلى نسبة من بين الخمائر.

الكلمات المفتاحية: الفطريات، الرشاشيات، المبيضات ومسحات بيئية.